# SEPARATION OF EPOXY, HYDROXY, HALOHYDROXY AND KETO FATTY ACIDS AND DERIVATIVES BY THIN-LAYER CHROMATOGRAPHY\*

M. W. ROOMI, M. R. SUBBARAM AND K. T. ACHAYA Regional Research Laboratory, Hyderabad (India) (Received January 18th, 1966)

Separation by thin-layer chromatography (TLC) of oxygenated fatty acid derivatives has been reported by SUBBARAO *et al.*<sup>1,2</sup>, MORRIS<sup>3-5</sup>, SGOUTAS AND KUM-MEROW<sup>6</sup> and KAUFMANN AND KO<sup>7</sup>. The use of glycol complexing agents such as boric acid, sodium borate and sodium arsenite for the separation of *erythro* and *threo* isomers of di- and tetrahydroxy fatty acids was first reported by MORRIS<sup>3</sup>. In later work<sup>4, 5</sup>, he coupled this elegant technique with the use of silver nitrate as an ethenoid complexing agent<sup>3, 8, 9</sup> to separate methyl esters of various positional and geometrical isomers of oxygenated fatty acids differing in degree of unsaturation. In this paper, the TLC behaviour of a number of hydroxy, epoxy, halohydroxy and keto fatty acids, esters and alcohols prepared as earlier reported<sup>10–13</sup>, using direct (D), reversedphase (RP) and boric acid-coated (B) silica gel plates is described.

## MATERIALS AND METHODS

The various epoxy fatty acids, esters and alcohols used in this study were prepared according to the method of FINDLEY et al.<sup>14</sup>. Erythro and three acids were prepared by hydroxylation of cis unsaturated acids with alkaline potassium permanganate<sup>15</sup> and performic acid<sup>16</sup> and the esters were prepared by the usual procedure with methanol and sulphuric acid. Threo- and erythro-hydroxyacetoxy compounds were prepared by refluxing cis- and trans-epoxy compounds with acetic acid while the diacetoxy fatty acids were obtained by treating the dihydroxy acids with acetic anhydride and pyridine. Erythro- and threo-2,3-dihydroxydocosanoic acids were prepared by the method of ROOMI et al.<sup>13</sup>. Halohydroxy compounds were prepared according to the method of KING<sup>17</sup>. 9(10)-Hydroxyoctadecanoic and 13(14)-hydroxydocosanoic acids were obtained by catalytic hydrogenation of cis-9,10-epoxystearic and 13,14-epoxydocosanoic acids<sup>11</sup> and the 9(10)-ketostearic acid by chromium trioxide oxidation of the 9(10)-hydroxy acid<sup>18</sup>. Both the 9-hydroxy-10-keto and 10-hydroxy-9-ketostearic acids were prepared by neutral permanganate oxidation of oleic acid by KING's method<sup>19</sup>. Permanganate oxidation of 9,10-stearolic acid<sup>20</sup> gave the 9,10-diketostearic acid. Reduction of esters using lithium aluminium hydride in tetrahydrofuran gave the corresponding alcohols.

The direct TLC procedure in use in this laboratory, employing Desaga equipment,  $275-\mu$  layers of silica gel G, and ether-light petroleum for development, has

\* Taken in part from the Ph.D. Thesis of M. W. ROOMI submitted to the Osmania University in September, 1965. been described earlier<sup>1</sup>. For reversed-phase TLC, the dried, coated plate was uniformly impregnated with silicone oil (Dow Corning silicone fluid 200) by allowing a 5% solution in ether to ascend the plate in a developing chamber<sup>21</sup>. Both for acids and esters, only one solvent system was used for reversed-phase TLC, *viz*. acetonitrileacetic acid-water (70:10:20, v/v). Impregnation with boric acid (2.8 g/25 g silica gel) was carried out by the method of MORRIS<sup>4</sup>, and ether-light petroleum mixtures were used for development.

#### RESULTS AND DISCUSSION

# Epoxy compounds

The separation of epoxy acids, esters and alcohols by direct and reversedphase TLC is indicated in Table I. By DTLC *cis*-epoxy compounds are separated from their *trans* isomers. The monoepoxy has a higher mobility than the corresponding diepoxy compound. Positional isomers of the same carbon chain length are separable (*cis*-6,7- from *cis*-9,10-), and also epoxy compounds of different chain lengths (18 from 22). Esters move faster than either acids or alcohols. By RPTLC also, separations are possible on the basis both of chain length and the number of epoxy groups in the chain.

#### TABLE I

SEPARATION OF EPOXY ACIDS, ESTERS AND ALCOHOLS BY TLC

 $R_F$  values  $\times$  100.

D = direct, RP = reversed-phase.

| Compound                                   | Carbon                         | n Acids |    | Esters |                    | Alcohols          |    |
|--|--------------------------------|---------|----|--------|--------------------|-------------------|----|
|  | chain<br>length D <sup>n</sup> | RPb     | Da | RP     | $D^{\mathfrak{a}}$ | $RP^{\mathrm{b}}$ |    |
| cis-6,7-Epoxystearic acid                  | 18                             | 41      | 76 | 48     | 50                 | <sup>_</sup>      |    |
| rans-6,7-Epoxystearic acid                 | 18                             | 44      | 76 | 54     | 50                 |                   |    |
| cis-9,10-Epoxystearic acid                 | 18                             | 45      | 77 | 54     | 51                 | 30                | 71 |
| rans-9,10-Epoxystearic acid                | 18                             | 48      | 77 | 60     | 51                 | 34                | 70 |
| cis, cis-9, 10, 12, 13-Diepoxystearic acid | 18                             | 32      | ġĠ | 3      | 81                 | <u> </u>          |    |
| sis-13,14-Epoxydocosanoic acid             | 22                             | 48      | 64 | 72     | 33                 | 42                | 63 |
| vans-13,14-Epoxydocosanoic acid            | 22                             | 58      | 64 | So     | 33                 | 46                | 63 |

<sup>a</sup> Ether in light petroleum (30:70, v/v).

<sup>b</sup> Acetonitrile-acetic acid-water (70: 10: 20, v/v).

## Hydroxy acids, esters and alcohols

(a) Monohydroxy acids. By DTLC using 30 % ether in light petroleum, 2hydroxydocosanoic acid was separated from the 13(14)-hydroxy isomer (Table II). MORRIS<sup>5</sup> showed for hydroxystearic esters that two similar maxima occur for the 12(13)-hydroxy and the 2-hydroxy isomers respectively. In the present instance the 2-hydroxydocosanoic acid moves faster than the 13-hydroxy isomer ( $R_F \times 100, 96$ and 85 respectively). 6(7)- and 9(10)-Hydroxyoctadecanoic acids and esters are separable from each other, the 6-hydroxy isomer having a lower  $R_F$  value. Close positional isomers (6- and 7-, 9- and 10-hydroxy) can be well separated, confirming TABLE II

SEPARATION OF MONOHYDROXY ACIDS AND ESTERS BY TLC  $R_F$  values  $\times$  100. D = direct.

| Compounds  | Acids                      | Esters                       |
|--|----------------------------|------------------------------|
| · · · · · · · · · · · · · · · · · · ·  | Da                         | $D^{\mathrm{b}}$             |
| 6(7)-Hydroxyoctadecanoic acid<br>9(10)-Hydroxyoctadecanoic acid<br>2-Hydroxydocosanoic acid<br>13(14)-Hydroxydocosanoic acid | 62,57<br>71,66<br>96<br>85 | 27, 22<br>36, 30<br>93<br>78 |

<sup>n</sup> Ether-light petroleum (30:70, v/v).

<sup>b</sup> Ether-light petroleum (20:80, v/v).

an earlier report<sup>2</sup>. Separation is also effected according to chain length, e.g.  $C_{18}$  from  $C_{22}$ , the latter having greater mobility.

(b) Dihydroxy acids, esters and alcohols. Table III shows the separation of dihydroxy compounds by D-, RP- and BTLC. As with epoxy compounds, RPTLC separates according to carbon chain length and number of hydroxy groups, and is ineffective in separating stereoisomers. DTLC on the other hand shows slight separation of erythro and threo isomers. Incorporation of boric acid as now well established causes the threo isomer to move considerably faster than the erythro. Positional dihydroxy isomers (6,7- and 9,10-dihydroxyoctadecanoic acids and 2,3- and 13,14- dihydroxydocosanoic acids) are separable by DTLC and BTLC but not by RPTLC. The behaviour of 2,3-dihydroxydocosanoic acids is anomalous. By analogy with the

#### TABLE III

SEPARATION OF DI- AND TETRAHYDROXY ACIDS, ESTERS AND ALCOHOLS BY TLC  $R_F$  values  $\times$  100.

D = direct, B = boric acid-coated, RP = reversed-phase.

|  | Acids              |                    | Esters |                               |            | Alcohols |             |         |
|--|--------------------|--------------------|--------|-------------------------------|------------|----------|-------------|---------|
|  | $D^{\mathfrak{n}}$ | $B^{\mathfrak{b}}$ | RPc    | $\overline{D^{\mathfrak{a}}}$ | Bb         | RP       | Da          | $B^{d}$ |
| threo-6,7-Dihydroxystearic acid                | 8                  | 42                 | 84     | 24                            | 55         | 78       |             |         |
| erythro-6,7-Dihydroxystearic acid              | 5                  | 30                 | 84     | 20                            | 29         | 78       | <del></del> |         |
| threo-9,10-Dihydroxystearic acid               | 14                 | 48                 | 85     | 33                            | 60         | 78       |             |         |
| erythro-9,10-Dihydroxystearic acid             | 10                 | 35                 | 85     | 29                            | 33         | 78       |             | <u></u> |
| threo, threo-9, 10, 12, 13-Tetrahydroxystearic |                    |                    |        |                               |            |          |             |         |
| acid   | o                  | 10                 | 94     | Ο.                            | 17         | 87       | <u> </u>    | ·       |
| rythro, erythro-9, 10, 12, 13-Tetrahydroxy-    |                    |                    | •      |                               | •          | •••      |             |         |
| stearic acid                                   | 0                  | 5                  | 94     | ο                             | 10         | 87       | ·           | · · ·   |
| hreo-2,3-Dihydroxydocosanoic acid              | о                  | ō                  |        | 60                            | 64         |          | 23          | 45      |
| erythro-2,3-Dihydroxydocosanoic acid           | о                  | 0                  |        | 55                            | <b>5</b> Ġ |          | 22          | 44      |
| threo-13,14-Dihydroxydocosanoic acid           | 25                 | 61                 | 72     | 46                            | 84         | 64       | 55          | 53      |
| erythro-13,14-Dihydroxydocosanoic acid         | 22                 | 48                 | 72     | 39                            | 43         | 64       | 54          | 47      |

Ether-light petroleum (50:50, v/v).

<sup>b</sup> Ether-light petroleum (40:60, v/v).

<sup>c</sup> Acetonitrile-acetic acid-water (70:10:20, v/v).

<sup>d</sup> Ether-light petroleum (70:30, v/v).

2-monohydroxy acid, the 2,3-dihydroxy acids were expected to move faster than the corresponding 13,14-isomers. However, both the *erythro-* and *threo-2,3-dihydroxy-* docosanoic acids did not move from the starting line even when pure ether or methanol was used as solvent. As expected, however, esters of 2,3-dihydroxydocosanoic acids moved farther than the corresponding 13,14-isomers on DTLC. *Erythro* and *threo* isomers were well separated on a boric acid-coated plate. In this instance the 2,3-dihydroxy esters had a lower mobility than the 13,14-isomer.

The 1,2,3- and 1,13,14-docosanetriols were separable both by DTLC and BTLC. Using BTLC, the *erythro* and *threo* isomers of 1,13,14-docosanetriols were separated from each other but not the isomers of 1,2,3-docosanetriols.

A dihydroxy compound can be separated from a tetrahydroxy compound by all three methods. Ester separations were superior, but the actual resolution of acids recorded could be further improved by increasing the volume of ether in the solvent system.

Some aspects of separation of *erythro*- and *threo*-dihydroxy fatty acids and their esters have been discussed by MORRIS<sup>4</sup>. Similar separation of carbohydrates has been attributed to ease of dehydration to form a five-membered acidic complex<sup>22</sup>. When one or both hydroxy groups were acetylated, TLC separation on boric acid plates no longer occurred (Table IV) probably because of the absence of protons from vicinal hydroxy groups necessary for complex formation.

## TABLE IV

SEPARATION OF DIHYDROXY, HYDROXYACETOXY AND DIACETOXY ACIDS BY TLC  $R_F$  values  $\times$  100.

| Compound  | Da     | Ba     |
|---|--------|--------|
| threo-9,10-Dihydroxystearic acid                | 7      | 34     |
| erythro-9,10-Dihydroxystearic acid              | 3      | 28     |
| threo-9(10)-Hydroxy-10(9)-acetoxystearic acid   | 35, 26 | 43, 50 |
| erythro-9(10)-Hydroxy-10(9)-acetoxystearic acid | 35, 26 | 43, 50 |
| threo-9,10-Diacetoxystearic acid                | 52     | 74     |
| erythro-9,10-Diacetoxystearic acid              | 52     | 74     |

D = direct, B = boric acid-coated.

<sup>a</sup> Ether-light petroleum (30:70, v/v).

The diacetoxy acids moved faster than the others, followed by the hydroxyacetoxy and the dihydroxy acids considerably behind. On opening a 9,10-epoxy ring with acetic acid, the two isomers obtained, *viz.* 9-hydroxy-10-acetoxy and 9-acetoxy-10-hydroxy, are well separated by TLC, the spot intensity suggesting roughly equal proportions of the isomers, as during other ring openings of epoxy fatty acids<sup>11</sup>.

# Halohydroxy acids and esters

The separation of close positional isomers is also possible for halohydroxy acids (Table V). Separation both of positional isomers, e.g. 6,7- and 9,10-chloro-hydroxystearic acids and by chain length, e.g. 9,10-chlorohydroxystearic from 13,14- chlorohydroxydocosanoic acid is possible. Slight separation of even close positional isomers such as 9-chloro-10-hydroxy and 10-chloro-9-hydroxystearic acids is possible.

#### TABLE V

SEPARATION OF HALOHYDROXY ACIDS AND ESTERS BY TLC  $R_F$  values  $\times$  100. D = direct.

D = cifect.

| Compound                             | Acids              | Esters                        |  |
|--------------------------------------|--------------------|-------------------------------|--|
|                                      | $D^{\mathfrak{p}}$ | $\overline{D^{\mathfrak{a}}}$ |  |
| C <sub>18</sub> series               |                    | *                             |  |
| threo-6(7)-Chloro-7(6)-hydroxy       | 25, 22             | 29, 25                        |  |
| erythro-6(7)-Chloro-7(6)-hydroxy     | 25, 22             | 29, 25                        |  |
| threo-9(10)-Chloro-10(9)-hydroxy     | 31, 28             | 35, 31                        |  |
| erythro-9(10)-Chloro-10(9)-hydroxy   | 32, 28             | 35, 31                        |  |
| C22 series                           |                    |                               |  |
| threo-13(14)-Chloro-14(13)-hydroxy   | 44                 | 67                            |  |
| erythro-13(14)-Chloro-14(13)-hydroxy | 40                 | 67                            |  |

<sup>a</sup> Ether-light petroleum (20:80, v/v).

Esters move faster than acids. Chloro-, bromo- and iodohydroxy acids are not separated from each other. RPTLC failed to give any of these separations except according to chain length.

# Hydroxy and keto acids

From Table VI a diketo acid is seen to move faster than a monoketo acid, which in turn moves faster than a monohydroxy acid. A monoketo acid has an  $R_F$ value between those of a diketo and a ketohydroxy acid, and similarly a monohydroxy acid lies between a dihydroxy and ketohydroxy acid. Positional ketohydroxy compounds are not separable. When the crude reaction product of peracid epoxidation of an unsaturated acid is separated by DTLC with 30 % ether-light petroleum, the unreacted unsaturated acid moves with the solvent front and the epoxy, dihydroxy and hydroxyacetoxy acids follow the pattern shown in Table VI.

# TABLE VI

SEPARATION OF EPOXY, HYDROXY AND KETO ACIDS BY TLC

 $R_F$  values  $\times$  100.

| D = | direct. |  |
|-----|---------|--|
|-----|---------|--|

| Compound                           | Acids  |
|------------------------------------|--------|
| •                                  | $D^n$  |
| cis-9,10-Epoxystearic acid         | 37     |
| 9(10)-Hydroxystearic acid          | 17, 13 |
| 9(10)-Ketostearic acid             | 37     |
| 9,10-Diketostearic acid            | 53     |
| 9-Hydroxy-10-ketostearic acid      | 21     |
| 10-Hydroxy-9-ketostearic acid      | 21     |
| threo-9,10-Dihydroxystearic acid   | 3      |
| erythro-9,10-Dihydroxystearic acid | I      |

<sup>B</sup> Ether-light petroleum (20:80, v/v).

97

The mechanism of separations obtained with various systems is thoroughly discussed by MORRIS<sup>5</sup>. Some further general conclusions may be drawn from the present experimental results. Compounds of the same carbon chain length carrying different groups like epoxy, mono-, di- and tetrahydroxy, keto, diketo and halohydroxy are separable from each other. Slight changes in the position of these groups in the carbon chain alter the polarity sufficiently to enable TLC separation. As is now well established, the use of boric acid-coated plates enables stereoisomers of di- and tetrahydroxystearic acids to be separated. Compounds of different carbon chain lengths can be separated by DTLC, BTLC and RPTLC, the last mentioned indeed effecting only this type of separation.

SUMMARY

Epoxy, halohydroxy, hydroxy, dihydroxy, keto, diketo and tetrahydroxy fatty acids of C<sub>18</sub> and C<sub>22</sub> chain lengths are separable by direct thin-layer chromatography. Erythro and threo di- and tetrahydroxy fatty acids are better separated on a boric acid-coated silica gel plate. Separation both of positional isomers (6,7 from 9,10 compounds) and of close positional isomers (9-hydroxy-10-acetoxy from 9acetoxy-10-hvdroxy) is possible. All the three systems, viz. direct, boric acid-coated and reversed-phase thin-layer chromatography, separate according to chain length The first two systems are generally considerably more useful for oxygenated products.

# REFERENCES

- 1 R. SUBBARAO, M. W. ROOMI, M. R. SUBBARAM AND K. T. ACHAYA, J. Chromatog., 9 (1962) 295.
- 2 R. SUBBARAO AND K. T. ACHAYA, J. Chromatog., 10 (1904) 235.
- 3 L. J. MORRIS, Chem. Ind. (London), (1962) 1238.

a ne nova co-dene so al avea a ndarijejska iz savanik novejska ce ujekteljske i

- 4 L. J. MORRIS, J. Chromatog., 12 (1903) 321.
- 5 L. J. MORRIS, J. Chromatog., 20 (1905) 27.
- 6 D. SGOUTAS AND F. A. KUMMEROW, J. Am. Oil Chemists' Soc., 40 (1963) 138.
- 7 H. P. KAUFMANN AND Y. S. KO, Fette, Seifen, Anstrichmittel, 63 (1961) 828.
- 8 B. DE VRIES, Chem. Ind. (London), (1962) 1049.
- 9 C. B. BARRETT, M. S. J. DALLAS AND F. B. PADLEY, Chem. Ind. (London) (1962) 1050.
- 10 M. W. ROOMI, M. R. SUBBARAM AND K. T. ACHAYA, Indian J. Chem., 1 (1963) 78.
- 11 M. W. ROOMI, M. R. SUBBARAM AND K. T. ACHAYA, Indian J. Chem., 3 (1965) 311. 12 M. W. ROOMI, M. R. SUBBARAM AND K. T. ACHAYA, personal communication.
- 13 M. W. ROOMI, M. R. SUBBARAM AND K. T. ACHAYA, personal communication
- 14 T. W. FINDLEY, D. SWERN AND J. T. SCANLAN, J. Am. Chem. Soc., 67 (1945) 412.
- 15 A. LAPWORTH AND E. N. MOTTRAM, J. Chem. Soc., (1925) 1628.
- 16 D. SWERN, G. N. BILLEN, T. W. FINDLEY AND J. T. SCANLAN, J. Am. Chem. Soc., 67 (1945) 1786.
- 17 G. KING, J. Chem. Soc., (1949) 1817.
- 18 C. H. MACK AND W. G. BICKFORD, J. Org. Chem., 18 (1953) 686.
- 19 G. KING, J. Chem. Soc., (1936) 1788.
- 20 N. A. KHAN AND M. S. NEWMAN, J. Org. Chem., 17 (1954) 1063. 21 M. W. ROOMI, M. R. SUBBARAM AND K. T. ACHAYA, J. Chromatog., 16 (1964) 106.
- 22 G. PASTUSKA, Z. Anal. Chem., 179 (1901) 427.

J. Chromatog., 24 (1966) 93-98